

Short sequence-paper

## The cDNA sequence of proton-pumping nicotinamide nucleotide transhydrogenase from man and mouse

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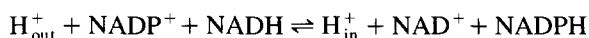
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### Abstract

cDNA clones for the human and mouse nicotinamide nucleotide transhydrogenases have been isolated and their sequences have been determined. Multiple alignments show that the functional proteins are encoded by single mRNAs. The deduced amino acid sequences are approximately 95% identical for the previously known bovine, and the human and mouse proteins. The major variable region is located in the presequence. It is proposed that all mammalian transhydrogenases have a similar structure.

**Keywords:** Transhydrogenase; Proton pumping; NAD; NADP; cDNA; (Human); (Mouse)

Nicotinamide nucleotide transhydrogenase (TH) (EC 1.6.1.1) is a redox-driven proton pump which catalyzes the reversible reduction of  $\text{NADP}^+$  by NADH according to the reaction



where protons are pumped from the cytosol or periplasmic space (out) to the matrix or the cytosol (in) in mitochondria and certain bacteria, respectively. Transhydrogenases are found in most microorganisms and all mammalian tissues investigated, and the transhydrogenase gene is also expressed in man [1]. The enzymes from bovine, *E. coli* and *Rhodospirillum rubrum* have been characterized extensively in their detergent-dispersed pure states as well as in reconstituted phospholipid vesicles [1]. The genes of these and several other transhydrogenases have been cloned and their cDNA/DNA sequenced [1], but only the *E. coli* [2,3] and *R. rubrum* [4] transhydrogenase genes have so far been overexpressed. Except for the bovine enzyme, which is composed of a homodimeric active form of a single subunit, other transhydrogenases are composed of two or three subunits also arranged as homodimeric active forms [1]. However, among the mammalian enzymes only the bovine transhydrogenase has been completely charac-

terized with respect to amino acid sequence [5]. It is therefore still uncertain whether the bovine enzyme is a general, single subunit, representative of mammalian transhydrogenases. In order to resolve this problem and to open the way for studies of transhydrogenase gene regulation in mammals, we have cloned the genes for the human and mouse transhydrogenases.

The cloning strategies were conducted essentially as described by Maniatis et al. [6]. Commercial cDNA libraries in the Lambda ZAP vector (936208 human male heart and 935302 mouse female liver, Stratagene, USA), were screened using cDNA from the bovine transhydrogenase as probe (construct pUGO4; Ref. [7]). The probe was <sup>32</sup>P-labeled with the Mega-Prime kit and <sup>32</sup>P-dATP obtained from Amersham (UK). Positive clones were identified by autoradiography. The libraries were screened as described by Stratagene. Several positive clones were isolated and clones with large inserts were chosen for sequencing. They were excised as pBluescript and single-stranded DNA rescues were performed as described by Stratagene. Clones were sequenced both as plasmid preparations (QIAGEN, USA) and as single-strand rescues. DNA sequencing was carried out by the dideoxynucleotide method of Sanger et al. [8] on an ALF (Automated Laser Fluorescent) DNA sequencer (Pharmacia, Sweden), using the AutoRead kit (Pharmacia, Sweden) and the Long Ranger, Hydrolink, gel (AT Biochem, USA). Both M13 reverse primer and internal fluorescein-labelled sequencing

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primers obtained from Scandinavian Gene Synthesis (Linköping, Sweden) were used.

The mouse sequence was covered by two overlapping clones and the human by three overlapping clones; all of these clones were completely sequenced. The sequences have been submitted to GenBank/EMBL: accession numbers Z49204 (mouse) and Z50101 (human).

As shown in Fig. 1 the human and mouse transhydrogenase cDNA sequences were translated and aligned to the bovine sequence with the GCG program PileUp [9]. (Fig. 1). The human and mouse proteins are identical to the bovine protein in length [5] and all have a 43 amino acid long pre-sequence [10]. Thus, all three proteins are single gene products. In contrast, the bacterial enzymes are 2–3 gene products. Like the mammalian enzymes, the parasite transhydrogenase genes are also single genes but the order

of transcription of the two halves of the genes is reversed [1]. The human and bovine proteins are 97% identical, whereas the mouse protein is approximately 94% identical to both the human and the bovine proteins. Differences are most abundant in the region 400–450 aa counted from the N-terminal of the mature protein. This hyper-variable region is located in the first predicted transmembrane  $\alpha$ -helix including the preceding and succeeding loops, according to a prediction model made for the bovine enzyme by Holmberg et al. [7].

The pre-sequences are less conserved, between 69% and 81% identical and mainly hydrophobic (Fig. 1). Alignment of the presequences as a helical wheel shows that there are four semi/conserved positive amino acids distributed mainly on one half of the wheel (Fig. 2). This pre-sequence probably directs the premature protein to the

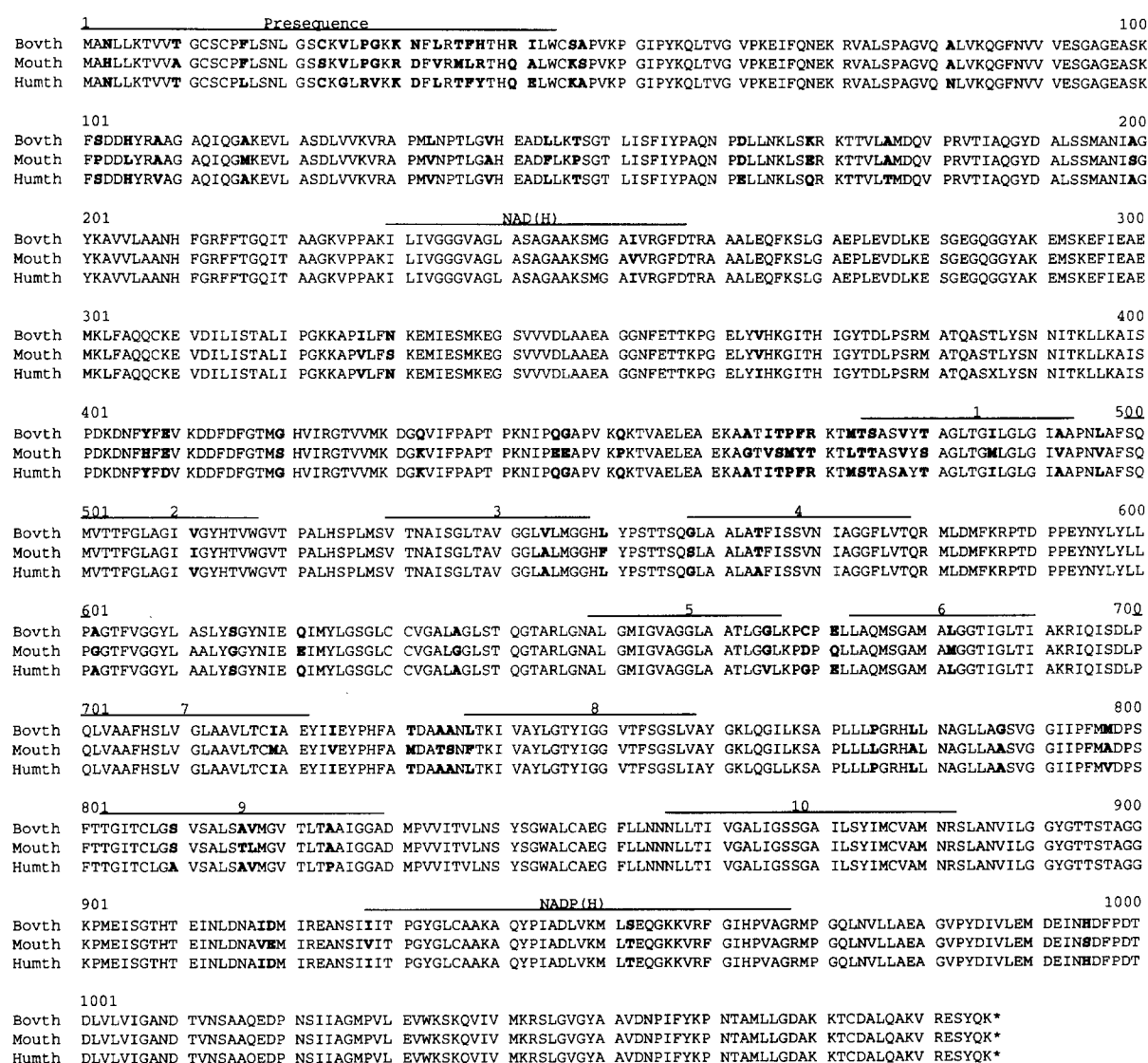


Fig. 1. Multiple alignment of the primary sequences for the bovine (Bovth), mouse (Mouth) and human (Humth) transhydrogenases. Predicted membrane spanning  $\alpha$ -helices are shown and numbered according to Holmberg et al. [7]. The proposed two substrate binding sites for NAD(H) and NADP(H), respectively, are also marked. Non-identical amino acids are bold. Stars denote stop codons.

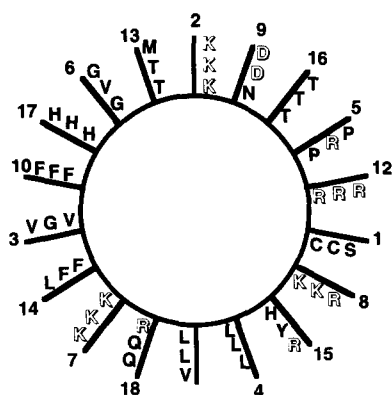


Fig. 2. Multiple alignment of the bovine, mouse and human transhydrogenase pre-sequences (aa 23–40) as a helical wheel. Uncharged residues are in plain text and charged residues are shadowed.

mitochondrion, where the charged surface may interact with a mitochondrial membrane receptor.

Based on the high similarity between the bovine, human and mouse transhydrogenases it is concluded that all mammalian transhydrogenases have a similar structure. The establishment of the human transhydrogenase sequence will enable more detailed studies on the regulation of the enzyme at the gene level and a possible correlation between expression and disease.

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